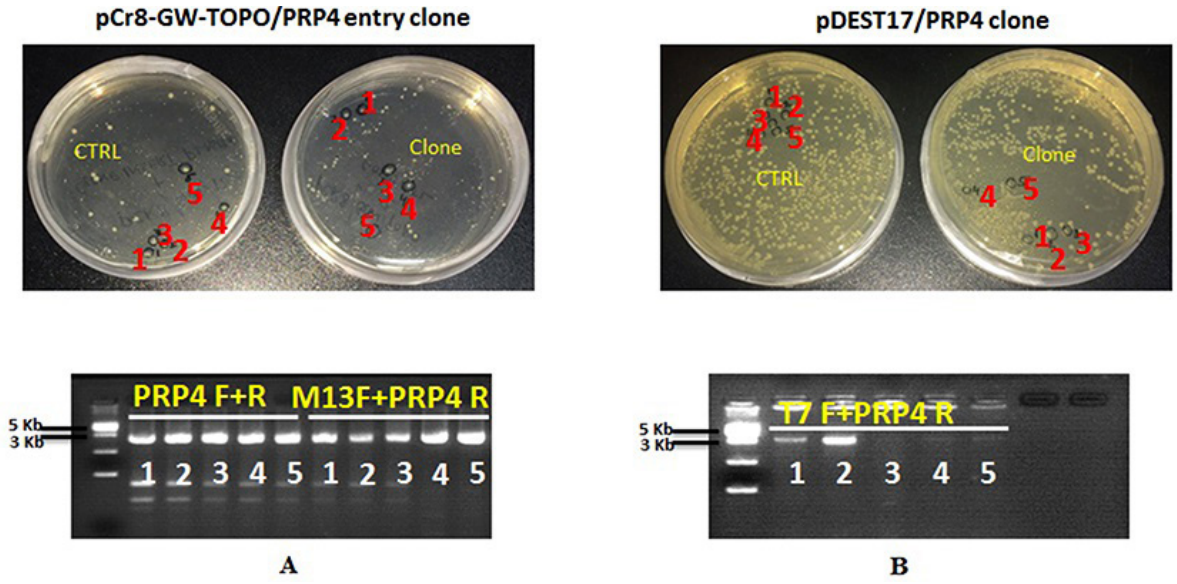
Supplementary Table S1. The PCR primers for *PRP4* and Gateway™ technology clones

PRP4 full-length gene (3024 bp)	
Forward	Reverse
PRP4 (1-21): ATGGCCGCCGCGGAGACCCA	PRP4 (3002-3024): TTAAATTTTTTCCTGGATGAAGG
PRP4 without the kinase domain (2058 bp)	
Forward	Reverse
PRP4 (1-21): ATGGCCGCCGCGGAGACCCA	PRP4 (2038-2059): ACGTTTATCTAGGACTTCACCTATGTTCAC
The pCr8-GW-TOPO/PRP4 entry clone (4875 bp)	
Forward	Reverse
PRP4 (1-21): ATGGCCGCCGCGGAGACCCA M13 Forward: GTAAAACGACGGCCAG	PRP4 (2038-2059): ACGTTTATCTAGGACTTCACCTATGTTCAC
The pDEST17/PRP4 clone (6784 bp)	
Forward	Reverse
T7 promoter: TAATACGACTCACTATAGG	PRP4 (2038-2059): ACGTTTATCTAGGACTTCACCTATGTTCAC



Supplementary Fig. S1. Confirmation of the construction of the pCr8-GW-TOPO/PRP4 entry and pDEST17/PRP4 clones. (A) The 2058 bp *PRP4* gene was cloned into the pCr8/GW/ TOPO vector to create an entry clone using the TOPO TA cloning kit according to the provided protocol. The pCr8/GW/ TOPO vector is resistant to spectinomycin. One Shot TOP10 *Escherichia coli* bacteria were used for the transformation. Five colonies were selected and cultured in LB medium with spectinomycin and the plasmids were obtained through miniprep. The confirmation of the entry clone was conducted through PCR using the specific primers given in Supplementary Table S1. The clones were further sequence-checked in the pCr8/GW/ TOPO vector and were as predicted (data not shown). (B) Entry clones from individual bacterial colonies were subsequently used in the LR reactions with the T7 promoter expression vector pDEST17. The figure demonstrates the *E. coli* colonies on an ampicillin-resistant LB agar plate. Confirmation of the pDEST17/PRP4 clone was conducted through PCR using the specific primers given in Supplementary Table S1. The clones were further sequence-checked in the pDEST17 vector and were as predicted (data not shown).